Evaluating immune correlates of protection in vaccine efficacy trials with stochastic-interventional causal effects

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Prelude:
Immune correlates of protection

The fights against HIV and COVID-19

- The HIV epidemic:
 - 1.5 million new infections occurring annually worldwide;
 - new infections outpace patients starting antiretroviral therapy;
 - HIV Vaccine Trials Network's (HVTN) 505 trial evaluated a novel antibody boost vaccine (Hammer et al. 2013).
- The COVID-19 epi pan endemic (Antia and Halloran 2021):
 - 270 331 619 643 686 772 million total cases globally (WHO);
 - new variants emerging, with vaccine uptake globally stalled;
 - COVID-19 Prevention Network's (CoVPN) COVE trial focused on Moderna's (mRNA-1273) vaccine (Baden et al. 2021).

Evaluating vaccine protection for HIV and COVID-19

- In 505: How would HIV infection risk have differed had the boost vaccine modulated antibody responses differently?
- In COVE: How would COVID-19 disease risk have differed for alternative vaccine-induced immunogenic response profiles?
- Question: Can [HIV-1, COVID-19] vaccines be improved by the modulation of immunogenic response profiles? How?

Why measure and analyze immune correlates?

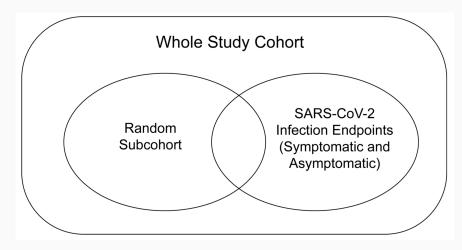
- Two interrelated goals of immune correlates analyses are to
 - identify/validate possible surrogate endpoints (Prentice 1989);
 - understand/delineate *protective mechanisms* of vaccination.
- If an immune correlate is established to reliably predict VE, subsequent efficacy trials may use it as a primary endpoint.
- Such surrogate endpoints may accelerate approval of
 - existing vaccines in different populations (e.g., in children);
 - new vaccines within the same class (e.g., based on mRNA);
 - inform the development of future vaccines (e.g., Ab targets);
 - shed light on immunologic mechanisms of disease occurrence.

Measuring correlates: Two-phase designs

- Often, use case-cohort design (Prentice 1986), a special case of two-phase sampling (Breslow et al. 2003).
- Phase 1: measure baseline, vaccination, endpoint on everyone.
- Phase 2: given baseline, vaccine, endpoint, select members of immune response subcohort with (possibly known) probability.
 - In 505: phase-two sample with 100% of HIV-1 cases and matching of non-cases (n = 189 per Janes et al. 2017)).
 - In *COVE*: stratified random subcohort ($n \approx 1600$) and all SARS-CoV-2 infection and COVID-19 disease endpoints.

A simple two-phase design: Case-cohort

Assaying over 30000 samples is expensive, statistically unnecessary.



Case-cohort design, per Prentice (1986), as applied to COVE.

Two-phase sampling masks the complete data structure

- Complete unobserved data $X = (L, A, S, Y) \sim P_0^X \in \mathcal{M}^X$:
 - L (baseline covariates): sex, age, BMI, behavioral HIV risk,
 - A (treatment): randomized assignment to vaccine/placebo,
 - S (exposure): immune response profile for relevant markers,
 - Y (outcome of interest): infection status at trial's end.
- Observed data $O = (B, BX) = (L, A, B, BS, Y) \sim P_0 \in \mathcal{M}$.
 - $B \in \{0,1\}$ indicates inclusion in the phase-two sample.
 - $\pi_0 := \mathbb{P}(B=1 \mid Y, L)$ must be *known by design* or estimated.

Act I:
Causal effects for quantitative exposures

Static interventions: What are they good for?

 Describe the manner in which X is hypothetically generated via an NPSEM-IE (Pearl 2009):

$$L = f_L(U_L); A \sim \text{Bern}(0.5); S = f_S(A, L, U_S); Y = f_Y(S, A, L, U_Y)$$

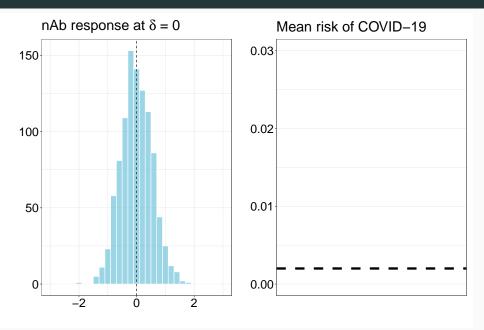
- Implies a model for distribution of counterfactual random variables induced by interventions on the system under study.
- A static intervention replaces (by "graph surgery") f_S with a specific value S = s in the conditional support $S \mid A = a, L$.
- This requires specifying a priori a particular value of exposure under which to evaluate the outcome — but what value?

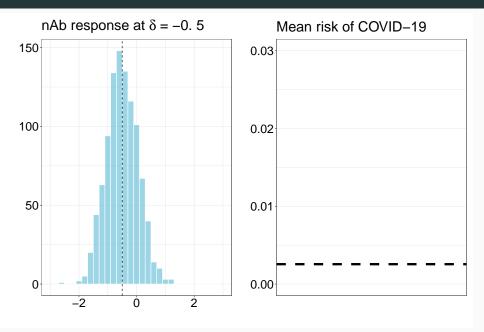
Controlled vaccine efficacy

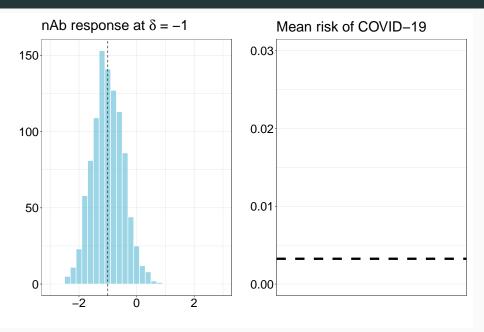
- For a hypothetical value $s \in \mathcal{S}$, the controlled direct effect (CDE) quantifies the effect of A on Y while fixing S = s.
- The hypothetical value S = s must be chosen carefully to be scientifically informative and to avoid positivity violations.
- For two hypothetical values $s_0, s_1 \in S$, Controlled Vaccine Efficacy (CVE) (Gilbert et al. 2022) is

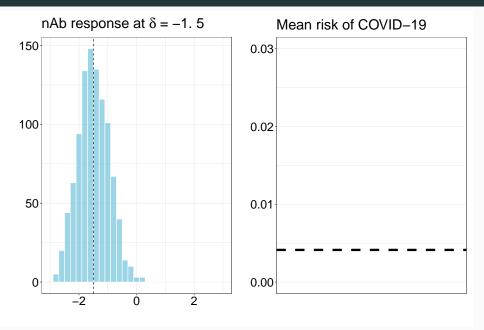
$$\mathsf{CVE}(s_0, s_1) = 1 - \frac{\mathbb{E}[\mathbb{P}(Y = 1 \mid S = s_1, A = 1, L = l)]}{\mathbb{E}[\mathbb{P}(Y = 1 \mid S = s_0, A = 0, L = l)]},$$

contrasting risk for vaccine receipt and $S = s_1$ vs. placebo and $S = s_0$, where $s_0 = 0$ is a plausible simplifying assumption in pathogen-naïve populations (Gilbert et al. 2024).









Stochastic interventions and modified treatment policies

- Stochastic interventions modify the value *S* would naturally assume by *modifying* the "natural" exposure distribution.
- Díaz and van der Laan (2012; 2018)'s shift interventions¹

$$d(s, l) = \begin{cases} s + \delta, & s + \delta < u(l) & \text{(if plausible)} \\ s, & s + \delta \ge u(l) & \text{(otherwise)} \end{cases}$$

• Causal estimand $\psi_{0,\delta}\coloneqq \mathbb{E}\{Y^{d(S,L)}\}$ is identified by

$$\psi_{0,\delta} = \int_{\mathcal{L}} \int_{\mathcal{S}} \mathbb{E}_{P_0} \{ Y \mid S = d(s, l), L = l \}$$
$$g_{0,S}(s \mid L = l) q_{0,L}(l) d\mu(s) d\nu(l)$$

 $^{^{1}}$ Haneuse and Rotnitzky (2013) introduced *modified treatment policies* (MTPs), extended by Díaz and van der Laan (2018) and Díaz et al. (2021).

Causally interpreting the statistical target parameter

Assumption 1: Stable Unit Treatment Value (SUTVA)

- $Y_i^{d(s_i,l_i)}$ does not depend on $d(s_j,l_j)$ for $i=1,\ldots,n$ and $j \neq i$, or lack of interference (Cox 1958)
- $Y^{d(s,l)} = Y$ in the event S = d(s,l)

Assumption 2: No Unmeasured Confounding

$$Y^{d(s,l)} \perp S \mid L = l$$

Assumption 3: Structural positivity

 $s \in \mathcal{S} \implies d(s, l) \in \mathcal{S}$ for all $l \in \mathcal{L}$, where \mathcal{S} denotes the support of S conditional on L = l

Stochastic-interventional vaccine efficacy (SVE)

Stochastic-interventional vaccine efficacy (SVE) estimand:

$$\begin{aligned} \mathsf{SVE}(\delta) &= 1 - \frac{\mathbb{E}[\mathbb{P}(\mathit{Y} = 1 \mid \mathit{S} = \mathit{d}(\mathit{s}, \mathit{l}), \mathit{A} = 1, \mathit{L} = \mathit{l})]}{\mathbb{P}(\mathit{Y}(0) = 1)} \\ &= 1 - \frac{\psi_{0,\delta}}{\mathbb{P}(\mathit{Y}(0) = 1)} \end{aligned}$$

- $\mathbb{P}(Y(0) = 1)$: counterfactual infection risk in the placebo arm under randomization, $\mathbb{P}(Y(0) = 1) \equiv \mathbb{P}(Y = 1 \mid A = 0)$.
- Summarizes VE as shifts of S by δ , yielding a VE measure based on hypothetically modifying S in the vaccine arm.

²CoVPN SAP: https://doi.org/10.6084/m9.figshare.13198595.

Estimation of the counterfactual mean $\psi_{0,\delta}$

A RAL estimator $\psi_{n,\delta}$ of $\psi_{0,\delta} \coloneqq \Psi(P_0)$ is asymptotically efficient if

$$\psi_{n,\delta} - \psi_{0,\delta} = \frac{1}{n} \sum_{i=1}^{n} D^{*}(P_{0})(O_{i}) + o_{P}(n^{-1/2}),$$

where $D^*(P)$ is the efficient influence function (EIF) of $\psi_{0,\delta}$ with respect to the nonparametric model \mathcal{M} at some distribution P.

The EIF of $\psi_{0,\delta}$ is indexed by two key *nuisance parameters*

$$\overline{Q}_Y(S,L) := \mathbb{E}_P(Y \mid S,L)$$
 conditional outcome mean $g_S(S \mid L) := f_P(S \mid L)$ generalized propensity score

Ensemble machine learning to estimate $\overline{Q}_{Y}(S, L)$ and $g_{S}(S \mid L)$.

Flexible, efficient, and doubly robust estimation

• Efficient influence function (EIF) of $\psi_{0,\delta}$ with respect to ${\mathcal M}$ is

$$D^{\star}(P_0)(o) = \frac{g_{0,S}(d^{-1}(s,l)|l)}{g_{0,S}(s|l)}(y - \overline{Q}_{0,Y}(s,l)) + \overline{Q}_{0,Y}(d(s,l),l) - \psi_{0,\delta}.$$

One-step estimator performs additive bias-correction:

$$\psi_n^+ = \frac{1}{n} \sum_{i=1}^n \overline{Q}_{n,Y}(d(S_i, L_i), L_i) + D_n^*(O_i) .$$

■ TML estimator tilts initial estimator $\overline{Q}_{n,Y}$ for bias-correction:

$$\psi_n^{\star} = \frac{1}{n} \sum_{i=1}^n \overline{Q}_{n,Y}^{\star}(d(S_i, L_i), L_i) .$$

■ *Doubly robust*: Consistent even if $\overline{Q}_{n,Y}$ or $g_{n,S}$ incorrect.

Act II:

Revenge of the two-phase sampling design ("There ain't nothing in this world for free")

Augmented Estimators for Two-Phase Sampling Designs

 Rose and van der Laan (2011) suggested inverse probability of censoring weighted (IPCW) loss functions:

$$\mathcal{L}(P_0^X)(O) = \frac{B}{\pi_0(Y, L)} \mathcal{L}(P_0^X)(X)$$

- When the sampling mechanism $\pi_0(Y, L)$ is known by design, this procedure yields a reasonably reliable estimator.
- When data-adaptive regression must be used that is, when $\pi_0(Y, L)$ is not known by design²— this is insufficient.

³Sampling of non-cases in HVTN 505 used matching (Janes et al. 2017).

Efficiency and Multiple Robustness (Hejazi et al. 2021)

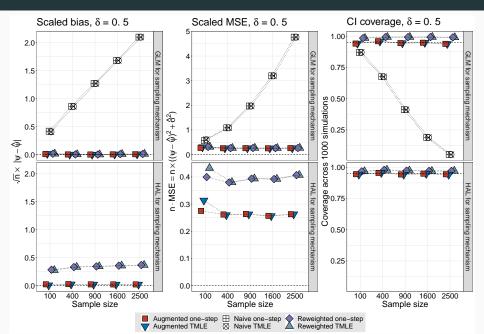
• Then, the IPCW augmentation must be applied to the EIF³:

$$D^{*}(P_{0}^{X})(o) = \frac{b}{\pi_{0}(y, l)} D_{F}^{*}(P_{0}^{X})(x) - \left(1 - \frac{b}{\pi_{0}(y, l)}\right)$$
$$\mathbb{E}(D_{F}^{*}(P_{0}^{X})(x) \mid B = 1, Y = y, L = l).$$

- Expresses observed data EIF $D^*(P_0^X)(o)$ via complete data EIF $D_F^*(P_0^X)(x)$; inclusion of second term improves efficiency.
- An emergent multiple robustness property combinations of $\{g_0(S \mid L), \overline{Q}_0(S, L)\} \times \{\pi_0(Y, L), \mathbb{E}(D_F^*(P_0^X)(x) \mid B = 1, Y, L)\}.$
- ullet Our txshift R package implements our estimators of $\psi_{0,\delta}.$

⁴Robins et al. (1994) explored a similar correction for such designs.

Comparing Reweighted and Augmented Estimators



Act III: Prediction and immunobridging via SVE

Prediction of HIV-1 risk (Hejazi et al. 2021)

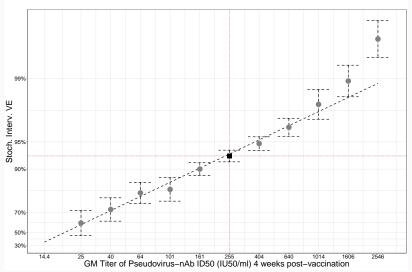
TML estimates of mean counterfactual HIV-1 infection risk under shifted CD8+ polyfunctionality with pointwise confidence intervals and summarization via working marginal structural model ($\hat{\beta}_{TMIF} = -0.013$) 0.10 0.08 ----Risk of HIV–1 infection 0.0 0.02 0.00 -2

HIV-1 risk change across CD8+ score (txshift R package).

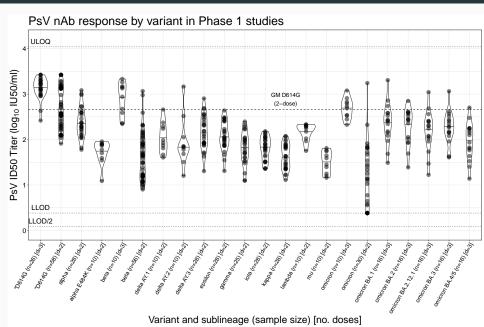
Posited change in standardized CD8+ polyfunctionality (sd units)

SVE prediction of mRNA-1273 VE (Huang et al. 2023)

Stoch. Interv. VE vs. COVID-19 (4 weeks post-vaccination with 100 days follow-up)



Pooled phase 1 studies: PsV nAb responses across variants



SVE bridging of mRNA-1273 VE (Hejazi et al. 2023)

Stoch. Interv. VE vs. COVID-19 (4 weeks post-vaccination with 100 days follow-up) After 2 doses of mRNA-1273 99% Stoch. Interv. VE 70% 50% 30% 10% GM Titer of Pseudovirus-nAb ID50 (IU50/ml) 4 weeks post-vaccination SARS-CoV-2 variant delta lambda

Summary of SVE prediction for immunobridging

- SVE prediction shows sharp changes in VE with shifts to the GM titer of the PsV nAb correlate in vaccinees.
- Bridging VE across variants indicates VE drops but stabilizes at 50%, if the model based on ancestral D614G strain holds.
- Post-2nd dose: For most variants (excepting omicron), the VE estimate ranges from 50% (mu) to about 80% (lambda).
- SVE predictions and real-world VE estimates well-correlated, but SVE predictions may be underestimates, as PsV nAb correlate is an *imperfect* causal *mediator* of total VE.

The big picture

- Flexible, realistic interventions can be used to formulate modified treatment policies (based on "natural" exposure).
- Modified treatment policies address causal questions about hypothetical manipulations of quantitative variables.
- Efficient estimators with double/multiple robustness can safely answer such questions while incorporating machine learning.
- Applying machine learning with causal inference yields robust predictions of VE and evidence for immunobridging.
- Open source software for such statistical analyses is critical for the methods to have any impact on real-world studies.

Thank you

Thanks for your attention! Any questions?

- ★ https://nimahejazi.org
- O https://github.com/nhejazi
- https://twitter.com/nshejazi
- Biometrics: https://doi.org/10.1111/biom.13375
- IJID: https://doi.org/10.1016/j.ijid.2023.09.012
- Viruses: https://doi.org/10.3390/v15102029
- Vaccine: https://doi.org/10.1016/j.vaccine.2024.02.071

Appendix

Immune Correlates of Protection (Plotkin and Gilbert 2012)

- Correlate of Protection (CoP): immune marker statistically predictive of vaccine efficacy, not necessarily mechanistic.
- Mechanistic CoP (mCoP): immune marker that is causally and mechanistically responsible for protection.
- Nonmechanistic CoP (nCoP): immune marker that is predictive but not a causal agent of protection.
- A CoP is a candidate surrogate endpoint (Prentice 1989) primary endpoint in future trials if reliably predictive.

Literature: Díaz and van der Laan (2012; 2018)

- Proposal: Evaluate outcome under an altered intervention distribution e.g., $P_{\delta}(g_{0,S})(S=s\mid L)=g_{0,S}(s-\delta(L)\mid L)$.
- Identification conditions for a statistical parameter of the counterfactual outcome $\psi_{0,\delta}$ under such an intervention.
- Show that the causal quantity of interest $\mathbb{E}_{P_0^{\delta}}\{Y_{d(S,L)}\}$ is identified by a functional of the distribution of O, i.e.,

$$\psi_{0,\delta} = \int_{\mathcal{L}} \int_{\mathcal{S}} \mathbb{E}_{P_0} \{ Y \mid S = d(s, l), L = l \}$$
$$g_{0,S}(s \mid L = l) \cdot q_{0,L}(l) d\mu(s) d\nu(l)$$

Literature: Haneuse and Rotnitzky (2013)

- Proposal: Characterization of stochastic interventions as modified treatment policies (MTPs).
- Assumption of piecewise smooth invertibility allows for the post-intervention distribution of any MTP to be recovered:

$$g_{0,S}(s \mid l; \delta) = \sum_{j=1}^{J(l)} \mathbb{I}_{\delta,j}\{h_j(s, l), l\}g_0\{h_j(s, l) \mid l\}h_j'(s, l)$$

 MTPs account for the natural value of exposure S yet may be interpreted as imposing an altered intervention mechanism.

Literature: Young et al. (2014)

- Establishes equivalence between g-formula when proposed intervention depends on natural value and when it does not.
- This equivalence leads to a sufficient positivity condition for estimating the counterfactual mean under MTPs via the same statistical functional studied in Díaz and van der Laan (2012).
- Extends earlier identification results, providing a way to use the same statistical functional to assess $\mathbb{E}Y_{d(S,L)}$ or $\mathbb{E}Y_{d(L)}$.
- The authors also consider limits on implementing shifts d(S, L), and address working in a longitudinal setting.

Flexible conditional density estimation of $g_{0,S}$

Díaz and van der Laan (2011)'s conditional density estimator:

$$g_{n,\alpha}(s \mid L) = \frac{\mathbb{P}(s \in [\alpha_{t-1}, \alpha_t) \mid L)}{\alpha_t - \alpha_{t-1}}.$$

- Re-expressed as hazard regressions in repeated measures data.
- Tuning parameter $t \approx \text{bandwidth in kernel density estimation}$.
- When càdlàg (RCLL) with finite sectional variation, we have

$$\operatorname{logit}\{\mathbb{P}(s \in [\alpha_{t-1}, \alpha_t) \mid L)\} = \beta_0 + \sum_{w \subset \{1, \dots, d\}} \sum_{i=1}^n \beta_{w,i} \phi_{w,i},$$

for appropriate basis functions $\{\phi_{w,i}\}_{i=1}^n$ (Gill et al. 1995).

Flexible conditional density estimation of $g_{0,S}$

- Utilizing a particular basis construction for ϕ_w , van der Laan (2017)'s HAL estimator achieves $n^{-1/4}$ convergence rate⁴.
- Loss-based cross-validation allows selection of a suitable HAL estimator, which has only the ℓ_1 regularization term λ :

$$\beta_{n,\lambda} = \mathop{\arg\min}_{\beta: |\beta_0| + \sum_{w \subset \{1, \dots, d\}} \sum_{i=1}^n |\beta_{w,i}| < \lambda} P_n \mathcal{L}(g_{\beta,\lambda,S}),$$

where $\mathcal{L}(\cdot)$ is an appropriate loss function, giving $\{\lambda_n, \beta_n\}$.

- We denote by $g_{n,\lambda,S} \coloneqq g_{\beta_{n,\lambda},S}$, the HAL estimate of $g_{0,S}$.
- Our haldensify R package implements our estimator of $g_{0,S}$.

 $^{^6}$ Similar rates can be achieved via *local* (vs. global) smoothness assumptions on $g_{n,S}$ (see, e.g., Robins et al. 2008, Mukherjee et al. 2017, Liu et al. 2021).

Algorithm for TML estimation

- 1. Construct initial estimators g_n of $g_0(S, L)$ and Q_n of $\overline{Q}_0(S, L)$, perhaps using data-adaptive regression techniques.
- 2. For each observation i, compute an estimate $H_n(s_i, l_i)$ of the auxiliary covariate $H(s_i, l_i)$.
- 3. Estimate the parameter ϵ in the logistic regression model

$$logit\overline{Q}_{\epsilon,n}(s,l) = logit\overline{Q}_n(s,l) + \epsilon H_n(s,l),$$

or an alternative regression model incorporating weights.

4. Compute TML estimator Ψ_n of the target parameter, defining update \overline{Q}_n^* of the initial estimate $\overline{Q}_{n,\epsilon_n}$:

$$\Psi_n = \Psi(P_n^*) = \frac{1}{n} \sum_{i=1}^n \overline{Q}_n^*(d(S_i, L_i), L_i).$$

Algorithm for IPCW-TML estimation

- 1. Using all observed units (X), estimate sampling mechanism $\pi(Y, L)$, perhaps using data-adaptive regression methods.
- 2. Using only observed units in the phase-two sample B=1, construct initial estimators $g_n(S,L)$ and $\overline{Q}_n(S,L)$, weighting by the sampling mechanism estimate $\pi_n(Y,L)$.
- 3. With the approach described for the full data case, compute $H_n(s_i, l_i)$, and fluctuate submodel via logistic regression.
- 4. Compute IPCW-TML estimator Ψ_n of the target parameter, by solving the IPCW-augmented EIF estimating equation.
- 5. Iteratively update estimated sampling weights $\pi_n(Y, L)$ and IPCW-augmented EIF, updating TMLE in each iteration.

References

- Antia, R. and Halloran, M. E. (2021). Transition to endemicity: Understanding covid-19. Immunity, 54(10):2172–2176.
- Baden, L. R., El Sahly, H. M., Essink, B., Kotloff, K., Frey, S., Novak, R., Diemert, D., Spector, S. A., Rouphael, N., Creech, C. B., McGettigan, J., Khetan, S., Segall, N., Solis, J., Brosz, A., Fierro, C., Schwartz, H., Neuzil, K., Corey, L., Gilbert, P., Janes, H., Follmann, D., Marovich, M., Mascola, J., Polakowski, L., Ledgerwood, J., Graham, B. S., Bennett, H., Pajon, R., Knightly, C., Leav, B., Deng, W., Zhou, H., Han, S., Ivarsson, M., Miller, J., Zaks, T., and the COVE Study Group (2021). Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. New England Journal of Medicine, 384(5):403–416.
- Breslow, N., McNeney, B., Wellner, J. A., et al. (2003). Large sample theory for semiparametric regression models with two-phase, outcome dependent sampling. The Annals of Statistics, 31(4):1110–1139.
- Cox, D. R. (1958). Planning of Experiments. Wiley.
- Díaz, I. and van der Laan, M. J. (2011). Super learner based conditional density estimation with application to marginal structural models. *International Journal of Biostatistics*, 7(1):1–20.
- Díaz, I. and van der Laan, M. J. (2012). Population intervention causal effects based on stochastic interventions. Biometrics, 68(2):541–549.
- Díaz, I. and van der Laan, M. J. (2018). Stochastic treatment regimes. In Targeted Learning in Data Science: Causal Inference for Complex Longitudinal Studies, pages 167–180. Springer Science & Business Media.
- Díaz, I., Williams, N., Hoffman, K. L., and Schenck, E. J. (2021). Nonparametric causal effects based on longitudinal modified treatment policies. *Journal of the American Statistical Association*.
- Gilbert, P. B., Fong, Y., Hejazi, N. S., Kenny, A., Huang, Y., Carone, M., Benkeser, D., and Follmann, D. (2024). Four statistical frameworks for assessing an immune correlate of protection (surrogate endpoint) from a randomized, controlled, vaccine efficacy trial. *Vaccine*, 42(9):2181–2190.

- Gilbert, P. B., Fong, Y., Kenny, A., and Carone, M. (2022). A controlled effects approach to assessing immune correlates of protection. *Biostatistics*, (in press).
- Gill, R. D., van der Laan, M. J., and Wellner, J. A. (1995). Inefficient estimators of the bivariate survival function for three models. In *Annales de l'IHP Probabilités et statistiques*, volume 31, pages 545–597.
- Hammer, S. M., Sobieszczyk, M. E., Janes, H., Karuna, S. T., Mulligan, M. J., Grove, D., Koblin, B. A., Buchbinder, S. P., Keefer, M. C., Tomaras, G. D., et al. (2013). Efficacy trial of a DNA/rAd5 HIV-1 preventive vaccine. New England Journal of Medicine. 369(22):2083–2092.
- Haneuse, S. and Rotnitzky, A. (2013). Estimation of the effect of interventions that modify the received treatment. Statistics in Medicine, 32(30):5260–5277.
- Hejazi, N. S., Shen, X., Carpp, L. N., Benkeser, D., Follmann, D., Janes, H. E., Baden, L. R., El Sahly, H. M., Deng, W., Zhou, H., Leav, B., Montefiori, D. C., and Gilbert, P. B. (2023). Stochastic interventional correlates of protection analysis of the COVE trial, with application to predict mRNA-1273 vaccine efficacy against

SARS-CoV-2 variants. under review at Lancet Microbe.

15(10):2029.

- Hejazi, N. S., van der Laan, M. J., Janes, H. E., Gilbert, P. B., and Benkeser, D. C. (2021). Efficient nonparametric inference on the effects of stochastic interventions under two-phase sampling, with applications to vaccine efficacy trials. *Biometrics*.
- Huang, Y., Hejazi, N. S., Blette, B., Carpp, L. N., Benkeser, D., Montefiori, D. C., McDermott, A. B., Fong, Y., Janes, H. E., Deng, W., Zhou, H., Houchens, C. R., Martins, K. A., Jayashankar, L., Flach, B., Lin, B. C., O'Connell, S., McDanal, C., Eaton, A., Sarzotti-Kelsoe, M., Lu, Y., Yu, C., Kenny, A., Carone, M., Huynh, C.,
- O'Connell, S., McDanal, C., Eaton, A., Sarzotti-Kelsoe, M., Lu, Y., Yu, C., Kenny, A., Carone, M., Huynh, C., Miller, J., El Sahly, H. M., Baden, L. R., Jackson, L. A., Campbell, T. B., Clark, J. L., Andrasik, M. P., Kublin, J. G., Corey, L., Neuzil, K. M., Pajon, R., Follmann, D. A., Donis, R. O., Koup, R. A., Gilbert, P. B., and on behalf of the Immune Assays Team; Moderna, Inc., Team; Coronavirus Prevention Network (CoVPN)/Coronavirus Efficacy (COVE) Team; and the United States Government (USG)/CoVPN Biostatistics Teams (2023). Stochastic interventional vaccine efficacy and principal surrogate analyses of antibody markers as correlates of protection against symptomatic COVID-19 in the COVE mRNA-1273 trial. Viruses,

- Janes, H. E., Cohen, K. W., Frahm, N., De Rosa, S. C., Sanchez, B., Hural, J., Magaret, C. A., Karuna, S., Bentley, C., Gottardo, R., et al. (2017). Higher t-cell responses induced by dna/rad5 hiv-1 preventive vaccine are associated with lower hiv-1 infection risk in an efficacy trial. The Journal of Infectious Diseases.
- Liu, L., Mukherjee, R., Robins, J. M., and Tchetgen Tchetgen, E. (2021). Adaptive estimation of nonparametric functionals. *Journal of Machine Learning Research*, 22(99):1–66.
- Mukherjee, R., Newey, W. K., and Robins, J. M. (2017). Semiparametric efficient empirical higher order influence function estimators. arXiv preprint arXiv:1705.07577.
- Pearl, J. (2009). Causality: Models, Reasoning, and Inference. Cambridge University Press.

215(9):1376-1385.

Medicine, 8(4):431-440.

- Plotkin, S. A. and Gilbert, P. B. (2012). Nomenclature for immune correlates of protection after vaccination. Clinical Infectious Diseases. 54(11):1615–1617.
- Prentice, R. L. (1986). A case-cohort design for epidemiologic cohort studies and disease prevention trials. Biometrika, 73(1):1–11.
- Prentice, R. L. (1989). Surrogate endpoints in clinical trials: definition and operational criteria. Statistics in
- Robins, J. M., Li, L., Tchetgen Tchetgen, E., and van der Vaart, A. W. (2008). Higher order influence functions and minimax estimation of nonlinear functionals. In *Probability and statistics: essays in honor of David A.*
- Freedman, pages 335–421. Institute of Mathematical Statistics.

 Robins, J. M., Rotnitzky, A., and Zhao, L. P. (1994). Estimation of regression coefficients when some regressors
- are not always observed. *Journal of the American statistical Association*, 89(427):846–866.

 Rose, S. and van der Laan, M. J. (2011). A targeted maximum likelihood estimator for two-stage designs. *The*
- International Journal of Biostatistics, 7(1):1–21.
- van der Laan, M. J. (2017). A generally efficient targeted minimum loss based estimator based on the highly adaptive lasso. *International Journal of Biostatistics*, 13(2).
- Young, J. G., Hernán, M. A., and Robins, J. M. (2014). Identification, estimation and approximation of risk under interventions that depend on the natural value of treatment using observational data. *Epidemiologic Methods*, 3(1):1–19.